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Case report

Brincidofovir clearance of acyclovir-resistant herpes simplex virus-1 and adenovirus infection after stem cell transplantation

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Abstract:

Infections with adenovirus (AdV) and herpesviruses can result in considerable morbidity and mortality in pediatric hematopoietic stem cell transplant (SCT) recipients. Herpes simplex virus (HSV) reactivations are usually prevented by acyclovir (ACV) prophylaxis, whereas cidofovir (CDV) has been used off indication to manage AdV infections. We report a child with myelodysplastic syndrome undergoing multiple SCT, who experienced HSV-1 disease including severe mucositis and herpetic whitlow, as well as high viral load AdV DNAemia. Both ACV and CDV were ineffective; however, viral loads were decreased with brincidofovir, resulting in viral clearance. A subsequent Epstein–Barr virus disease with relevant meningoencephalitis responded to rituximab.

<u>Key words:</u> adenovirus; Epstein–Barr virus; herpes simplex virus; brincidofovir; hematopoietic stem cell transplantation

In healthy individuals, herpesvirus infections are usually controlled by the immune system, whereas they can lead to devastating conditions in immunosuppressed stem cell transplant (SCT) recipients. In these patients, herpes simplex virus type 1 (HSV-1) reactivations occur frequently without acyclovir (ACV) prophylaxis and occur as early as the pre-engraftment phase (1). Under ACV prophylaxis, HSV-1 reactivations in pediatric SCT recipients are rare and mainly caused by resistance-associated mutations in the viral thymidine kinase gene (2). If present, these mutations usually define cross-resistance to brivudine and the structurally similar nucleoside analogs penciclovir and its prodrug famciclovir (3). In addition, long-term ACV prophylaxis has been associated with a risk of developing ACV-resistant virus (4). The pyrophosphate analog foscarnet can be alternatively used for treatment, if no resistance-associated mutation is present in the viral DNA polymerase. B lymphocytes involved in lymphoproliferative disease occurring after SCT are targeted with rituximab.

Moreover, adenovirus (AdV) infections have been reported to contribute significantly to morbidity and mortality in pediatric SCT patients, especially during episodes of high level AdV DNAemia (5). Apart from a desirable but often challenging reduction of immunosuppression, cidofovir (CDV) has been proven useful for AdV DNAemia treatment but its use is frequently limited by nephro- and myelotoxicity (6). Brincidofovir (BCV) is an orally available analog of CDV containing a lipid side chain facilitating the transport of the molecule into the cell (7). Several groups have reported an inhibitory effect for double-stranded DNA viruses including AdV (8–12) and safety data in patients are published (7, 13). Analysis of a recent phase 3 trial to evaluate BCV

prophylaxis against cytomegalovirus demonstrated gastrointestinal adverse events and acute graft-versus-host disease was suspected and treated (14).

Here, we report a child with myelodysplastic syndrome (MDS) undergoing multiple SCT who experienced HSV-1 disease including severe mucositis and herpetic whitlow as well as high viral load AdV DNAemia. Both ACV and CDV were ineffective; however, viral loads were decreased with BCV, resulting in viral clearance.

Case report

A 5-year-old girl was diagnosed with MDS (FAB RC) and received a bone marrow graft from a 9/10 matched unrelated donor following conditioning according to the EWOG-MDS protocol (15). Both recipient and donor were initially HSV-1, Epstein–Barr virus (EBV), and AdV immunoglobulin-G seropositive. The day after SCT, the girl received standard ACV prophylaxis for prevention of HSV and varicella zoster virus infections and was continued until day 233 before foscarnet treatment commenced.

At 136 days after transplantation, the girl developed a secondary graft failure (see Fig. 1). She received a second reduced-intensity conditioning including fludarabine, alemtuzumab, cyclophosphamide, and melphalan with a subsequent, partially T-cell–depleted, peripheral blood SCT from the initial donor. On day 215 after initial SCT, she rejected again and received a third SCT from a new, also 9/10 identical matched-unrelated bone marrow donor after conditioning with fludarabine, thiotepa, 4 Gy total body irradiation, and anti-thymocyte globulin.

Despite ACV prophylaxis, HSV-1 reactivated and DNA became detectable in blood, cheek swab, as well as mouth washes after the second SCT. Phenotyping by

plaque-reduction assay revealed an ACV-resistant strain, and subsequent genotyping confirmed the presence of a C336Y resistance-associated mutation in the viral thymidine kinase gene (2). The girl presented with severe stomatitis and pharyngitis before herpetic whitlow developed on day 221 after the first SCT (Figs. 1 and 2A). Almost simultaneously, EBV DNAemia occurred and was treated with foscarnet (Fig. 1).

AdV DNA positivity in stool started 178 days and AdV DNAemia (species C) began 237 days after the first SCT, when second graft rejection occurred. CDV was started but AdV in plasma increased to >10⁶ copies/mL within 3 weeks, while HSV-1 DNAemia remained positive. Therefore, BCV was obtained by emergency use authorization and substituted for CDV and ACV. AdV and HSV-1 DNA in plasma became undetectable shortly after starting treatment. Herpetic whitlow resolved 75 days after BCV introduction (Figs. 1 and 2B).

An EBV rebound on day 286 with beginning lymphoproliferative disease developed into meningoencephalitis with neurologic deficits and was eventually treated with foscarnet and a single dose of rituximab.

Discussion

ACV has long been the mainstay in therapy for HSV diseases and is frequently used to prevent viral reactivation in the post-SCT period (9, 16). Although it is unusual, our patient reactivated HSV-1 under ACV prophylaxis and developed a fulminant herpetic whitlow. Hematologically, the situation was aggravated by recurrent episodes of graft rejection and constant lymphopenia, necessitating re-conditioning. This combination of graft rejections and multiple viral reactivations, including an ACV-resistant HSV-1 strain

and high viral load AdV DNAemia, led to a life-threatening condition. AdV treatment with CDV failed and required another approach with BCV, which has been shown to have *in vitro* activity against HSV-1 and AdV (17, 18).

In our patient, BCV controlled both HSV-1 and AdV DNAemia within a short period of time without adverse side effects, especially no impairment of hematopoiesis early after SCT. Still, a second EBV DNAemia occurred under BCV administration. Whether this reflects an inferior efficacy of BCV on EBV or simply the patient's reestablished lymphopoiesis remains unclear. However, the relevant EBV meningoencephalitis may indicate a poor penetration of BCV into the brain.

BCV is still being tested in clinical trials, and it is still too soon to provide a recommendation as to whether it might be useful in hematopoietic SCT recipients with viral infections (19). However, the need for novel compounds is obvious, and a beneficial effect of BCV in another pediatric HSCT recipient has been reported (20). Given the lack of available broad-spectrum antiviral agents and the frequent occurrence of multiple viral infections after SCT, BCV might prove a good option for these complicated patients.

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Conflict of interest: The authors declare that there is no conflict of interest.

References

- 1. Saral R, Burns WH, Laskin OL, Santos GW, Lietman PS. Acyclovir prophylaxis of herpes-simplex-virus infections. N Engl J Med 1981; 305 (2): 63–67.
- 2. Sauerbrei A, Bohn-Wippert K, Kaspar M, Krumbholz A, Karrasch M, Zell R. Database on natural polymorphisms and resistance-related non-synonymous mutations in thymidine kinase and DNA polymerase genes of herpes simplex virus type 1 and 2. J Antimicrob Chemother 2016; 71: 6–16.
- 3. Piret J, Boivin G. Resistance of herpes simplex viruses to nucleoside analogues: mechanisms, prevalence, and management. Antimicrob Agents Chemother 2011; 55 (2): 459–472.
- 4. Erard V, Wald A, Corey L, Leisenring WM, Boeckh M. Use of long-term suppressive acyclovir after hematopoietic stem-cell transplantation: impact on herpes simplex virus (HSV) disease and drug-resistant HSV disease. J Infect Dis 2007; 196 (2): 266–270.
- 5. Lion T. Adenovirus infections in immunocompetent and immunocompromised patients. Clin Microbiol Rev 2014; 27 (3): 441–462.
- 6. Sandkovsky U, Vargas L, Florescu DF. Adenovirus: current epidemiology and emerging approaches to prevention and treatment. Curr Infect Dis Rep 2014; 16 (8): 416.
- 7. Painter W, Robertson A, Trost LC, Godkin S, Lampert B, Painter G. First pharmacokinetic and safety study in humans of the novel lipid antiviral conjugate CMX001, a broad-spectrum oral drug active against double-stranded DNA viruses. Antimicrob Agents Chemother 2012; 56 (5): 2726–2734.

- 8. Florescu DF, Keck MA. Development of CMX001 (Brincidofovir) for the treatment of serious diseases or conditions caused by dsDNA viruses. Expert Rev Anti Infect Ther 2014; 12 (10): 1171–1178.
- 9. De SK, Hart JC, Breuer J. Herpes simplex virus and varicella zoster virus: recent advances in therapy. Curr Opin Infect Dis 2015; 28 (6): 589–595.
- Prichard MN, Kern ER, Hartline CB, Lanier ER, Quenelle DC. CMX001 potentiates the efficacy of acyclovir in herpes simplex virus infections.
 Antimicrob Agents Chemother 2011; 55 (10): 4728–4734.
- 11. Quenelle DC, Lampert B, Collins DJ, Rice TL, Painter GR, Kern ER. Efficacy of CMX001 against herpes simplex virus infections in mice and correlations with drug distribution studies. J Infect Dis 2010; 202 (10): 1492–1499.
- 12. Wold WS, Toth K. New drug on the horizon for treating adenovirus. Expert Opin Pharmacother 2015; 16 (14): 2095–2099.
- 13. Florescu DF, Pergam SA, Neely MN, et al. Safety and efficacy of CMX001 as salvage therapy for severe adenovirus infections in immunocompromised patients.

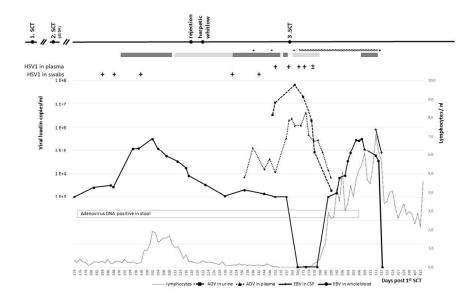
 Biol Blood Marrow Transplant 2012; 18 (5): 731–738.
- 14. Marty FM, Winston DJ, Chemaly RF, et al. Brincidofovir for prevention of cytomegalovirus (CMV) after allogeneic hematopoietic cell transplantation (HCT) in CMV-seropositive patients: a randomized, double-blind, placebocontrolled, parallel-group phase 3 trial. Biol Blood Marrow Transplant 2016; 22: S23.

- 15. Strahm B, Locatelli F, Bader P, et al. Reduced intensity conditioning in unrelated donor transplantation for refractory cytopenia in childhood. Bone Marrow Transplant 2007; 40 (4): 329–333.
- 16. Ito JF. Herpes simplex virus infections. In: Appelbaum FR, Forman SJ, Negrin RS, and Blume KG, eds. Thomas' Hematopoietic cell transplantation (4th edn). Chichester: Wiley-Blackwell, 2011: 3121–3136.
- 17. Florescu DF, Grimley MS, Bourne E, et al. Preliminary Data from 13 Liver Transplantation Patients who received Brincidofovir (BCV) for Adenovirus (AdV) Infection. Abstract, IDWeek, 2015.
- Vora SB, Brothers A, Englund JA. Renal Toxicity Outcomes for Pediatric Patients Receiving Cidofovir for the Treatment of Adenovirus Infections.
 Abstract, IDWeek, 2015.
- Matthes-Martin S, Boztug H, Lion T. Diagnosis and treatment of adenovirus infection in immunocompromised patients. Expert Rev Anti Infect Ther 2013; 11 (10): 1017–1028.
- 20. Paolino K, Sande J, Perez E, et al. Eradication of disseminated adenovirus infection in a pediatric hematopoietic stem cell transplantation recipient using the novel antiviral agent CMX001. J Clin Virol 2011; 50 (2): 167–170.

Figure legends

Fig. 1. Time course of virus reactivations and treatment. Viral loads of Epstein–Barr virus (EBV) measured in whole blood and cerebrospinal fluid, as well as of adenovirus (ADV) in ethylenediamene tetraacetic acid (EDTA) plasma and urine are shown in relation to the days after the first stem cell transplant (SCT). Polymerase chain reaction (PCR) followed standard protocols with a limit of detection of about 1000 copies/mL. Continued shedding of ADV in stool specimens was found between days 178 and 314 post SCT. Qualitative herpes simplex virus (HSV)-1 PCR was performed in EDTA plasma and oral fluids/swabs (days 184, 189 and 198), cheek swab (day 238), and digit swab (day 231). Resistance analyses, performed in the samples from days 189 (wild-type/mutant mixture) and 198 (mutant only), revealed the C336Y substitution in the viral thymidine kinase.

Fig. 2. (A) Herpetic whitlow before brincidofovir administration. (B) Herpetic whitlow after brincidofovir administration.



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Figure 2

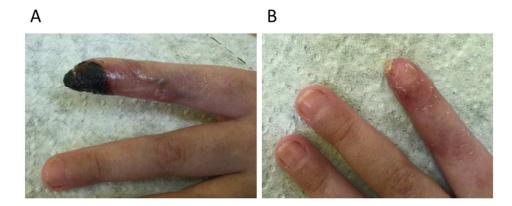


Figure 2 254x190mm (96 x 96 DPI)